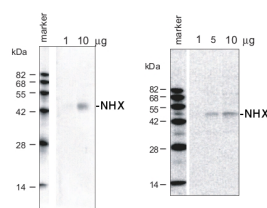


Product no **AS09 484****Na<sup>+</sup>/H<sup>+</sup> antiporter, sodium/hydrogen exchanger****Product information**

<b>Immunogen</b>	KLH-conjugated synthetic peptide derived from <i>Arabidopsis thaliana</i> NHX protein UniProt: <a href="#">Q68K14</a> , TAIR: <a href="#">At5g27150</a> ; chosen peptide is perfectly confirmed in AtNHX1 UniProt: <a href="#">Q0WVZ5</a> , partially in AtNHX2, UniProt: <a href="#">Q56XP4</a> , and not conserved in AtNHX3, UniProt: <a href="#">Q84WG1</a> and AtNHX4, UniProt: <a href="#">Q8S397</a> isoforms
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Immunogen affinity purified serum in PBS pH 7.4.
<b>Format</b>	Lyophilized
<b>Quantity</b>	50 µl
<b>Reconstitution</b>	For reconstitution add 50 µl of sterile water
<b>Storage</b>	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
<b>Additional information</b>	Protocol for vacuolar membrane isolation can be found <a href="#">here</a> .

**Application information**

<b>Recommended dilution</b>	1 : 8000 (ELISA), 1 : 1000 (WB)
<b>Expected   apparent MW</b>	59.5   45 kDa
<b>Confirmed reactivity</b>	<i>Arabidopsis thaliana</i> , <i>Kandelia obovata</i> , <i>Oryza sativa</i> , <i>Solanum lycopersicum</i> , <i>Vitis vinifera</i>
<b>Predicted reactivity</b>	<i>Gossypium hirsutum</i> , <i>Populus euphratica</i> , <i>Ricinus communis</i> , <i>Zea mays</i> (NHX1, NHX4) Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	Mangrove plants, <i>Nicotiana benthamiana</i> , sp. <i>Avicennia</i> ,
<b>Selected references</b>	<a href="#">Carmona-Salazar et al. (2021)</a> . Plasma and Vacuolar Membrane Sphingolipidomes: Composition and Insights on The Role of Main Molecular Species. <i>Plant Physiol.</i> 2021 Feb 11:kiab064. doi: 10.1093/plphys/kiab064. Epub ahead of print. PMID: 33570616. <a href="#">Cano-Ramirez et al. (2021)</a> M. Plasma Membrane Fluidity: An Environment Thermal Detector in Plants. <i>Cells.</i> 2021 Oct 17;10(10):2778. doi: 10.3390/cells10102778. PMID: 34685758; PMCID: PMC8535034. <a href="#">Prinsi et al. (2020)</a> . Root Proteomic Analysis of Two Grapevine Rootstock Genotypes Showing Different Susceptibility to Salt Stress. <i>Int J Mol Sci.</i> 2020 Feb 6;21(3). pii: E1076. doi: 10.3390/ijms21031076. <a href="#">Gupta and Shaw (2020)</a> . Biochemical and molecular characterisations of salt tolerance components in rice varieties tolerant and sensitive to NaCl: the relevance of Na <sup>+</sup> exclusion in salt tolerance in the species. <i>Funct Plant Biol.</i> 2020 Jul 30. doi: 10.1071/FP20089 <a href="#">Guo et al. (2018)</a> . Molecular Characterization of a Tonoplast Na <sup>+</sup> /H <sup>+</sup> Antiporter from Iris Lactea. Preprints 2018, 2018090557 (doi: 10.20944/preprints201809.0557.v1).

**Application example**

**1 µg and 10 µg of crude membrane fraction/lane** from *Arabidopsis thaliana* (left panel) and **1, 5 and 10 µg of crude membrane fraction/lane** *Raphanus sativus* L. (right panel) were separated on 12 % SDS-PAGE and blotted 1h to PVDF membrane (40 min. at 10 V using BioRad semidry transfer). Filters were blocked 1h with 5 % low-fat milk powder in TBS-T (0.05% Triton X.100). Membranes were washed 5 times with TBS-T, each time in a fresh polystyrene box and probed with anti-NHX antibodies (AS09 484, **1:1000**, 1h) and secondary anti-rabbit (**1:2000**, 1 h). Detection was done using chemiluminescence. All steps were performed in RT with agitation.