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Product no AS20 4416

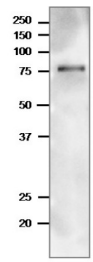
TGG1 | Myrosinase 1 (BGL38)

Product information

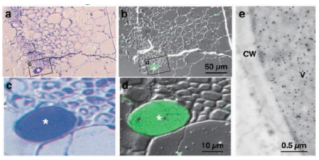
Immunogen	BSA-conjugated peptide, derived from N-terminus of *Arabidopsis thaliana* TGG1, UniProt: P37702, TAIR: At5g26000
Host	Rabbit
Clonality	Polyclonal
Purity	Total IgG. Protein A purified in PBS, 50% glycerol. Filter sterilized.
Format	Liquid at 2 mg/ml.
Quantity	200 µg
Storage	Store at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution	assay dependent (ELISA), 1: 1000 - 1: 2500 (IG), 1: 500 - 1: 1000 (IHC), 1: 1000-1: 3000 (WB)
Expected | apparent MW	61 | 77 kDa
Confirmed reactivity	*Arabidopsis thaliana*
Predicted reactivity	Species of your interest not listed? Contact us
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Additional information	19 amino acids of transit peptide are not present in the mature protein
Selected references	Farid et al. (2011). Arabidopsis thaliana alpha1,2-glucosyltransferase (ALG10) is required for efficient N-glycosylation and leaf growth. Plant J. 2011 Oct;68(2):314-25.doi: 10.1111/j.1365-313X.2011.04688.x. (Western blot) Shirakawa et al. (2010). Arabidopsis Qa-SNARE SYP2 proteins localized to different subcellular regions function redundantly in vacuolar protein sorting and plant development. Plant J. 2010 Dec;64(6):924-35.doi: 10.1111/j.1365-313X.2010.04394.x. (Western blot) Ueda et al. (2006). AtVAM3 is required for normal specification of idioblasts, myrosin cells. Plant Cell Physiol. 2006 Jan;47(1):164-75. doi: 10.1093/pcp/pci232. (Immunlocalization, Western blot).



Arabidopsis thaliana total leaf was freshly extracted with 2x SDS-sample buffer (+ 2ME) for SDS-PAGE and denatured with 4X SDS buffer at 95°C for 5 min. 10 µg of protein was loaded and separated on 15-20 % SDS-PAGE and blotted 1h to PVDF membrane. Blot was blocked with 3 % skim milk/TBS-T, 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 1000 in TBS-T for 1h/RT. The antibody solution was decanted and the blot was washed 4 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1h/RT with agitation. The blot was washed as above and developed with a chemiluminescent detection reagent, following manufacture's recommendations.



Immunolocalization of TGG1 in sections of *Arabidopsis thaliana* 48 days-old rosette leaves. Panel to the left: CBB staining (a,c). Middle panel: Reaction with anti-TGG1 antibodies used at 1: 1000 dilution and followed by visualization with AlexaFluor488 goat anti-rabbit antibodies at 1: 1000 (b.d). Right panel: electron microscopy of ultrathin sections mounted on Formvar-coated nickle grid. Anti-TGG1 antibodies were used at 1: 1000 dilution and following washing in PBS the sections were incubated with anti-rabbit IgG conjugated gold particles (AuroProbe EM). CW- cell wall, V-

vacuole.
Protocol:
Rosette leaves of *Arabidopsis thaliana* were fixed with 4% (w/v) paraformaldehyde and 1% glutaraldehyde in 0.05 M cacodylate buffer (pH 7.4) at 4°C for 3 h. After washing with 0.02 M cacodylate buffer (pH 7.4), these tissues were dehydrated with acetone and embedded in LR white resin at –20°C. Sections were cut on an ultramicrotome (Leica, Reichert Division, Vienna, Austria) for both light microscopic and electron microscopic analyses.